

## Cas9 Stable Cell Line

| Product Information             |                            |
|---------------------------------|----------------------------|
| Cell Line                       | HAP1 Cas9 Stable Cell Line |
| Parental (Horizon Discovery ID) | C631                       |
| Catalog ID                      | HD Cas9-011                |
| SNB ID                          | 42352                      |
| Cas9 expression promotor        | hEF1a                      |
| Passage                         | 20                         |
| Cryopreservation Date           | February 27, 2020          |

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| Total viable cells      | > 1x10 <sup>6</sup>          |
|-------------------------|------------------------------|
| Total Volume            | 1 mL                         |
| Cryopreservation Medium | 45% IMDM, 50% FBS, 5% DMSO   |
| Storage Conditions      | Liquid nitrogen vapour phase |

| Customer Support  |                                |
|-------------------|--------------------------------|
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| Quality Control  |  |        |
|------------------|--|--------|
| Test             | Test Method  | Result |
| Viability        | Post bank thawing and cultivation                          | Pass   |
| Sterility        | Direct inoculation of Tryptic Soy and Thioglycolate Broths | Pass   |
| Mycoplasma       | Mycoplasma detection by qPCR                               | Pass   |
| Characterisation | Functionality confirmed by gene editing assay (> 20%)      | Pass   |

| Growth Conditions                      |   |
|--|---|
| Recommended Culture Medium             | IMDM, 10% FBS, 1% Pen/Strep   |
| Cell Line Revival                      | Rapidly thaw cells in a 37°C water bath for 2 minutes until nearly (80%) thawed. Transfer contents into a tube containing pre-warmed media. Centrifuge the cells at 300 x g for 4 minutes and remove the supernatant. Add 2 mL of appropriate cell culture medium and transfer cells to T25 flask containing 4 mL of pre-warmed cell culture medium. Place cell in a humidified 37°C incubator with 5% CO <sub>2</sub> . Gently replace medium after 24 hours with 5-10 mL of appropriate cell culture medium and continue culturing at 37°C with 5% CO <sub>2</sub> . When appropriate (70-80% confluency), expand cell lines to a T75 flask using the subculturing procedures below.                        |
| Subculture                             | Carefully aspirate the growth medium from the cells. Gently wash cells with 7.5 mL PBS to remove the remaining media. Trypsinize the cells with 3 mL trypsin-EDTA solution. Place the flask in the 37 °C incubator for approximately 2 minutes or until the cells release from the flask. Add 15-30 mL of the appropriate Cell Culture Medium to resuspend the detached cells and inactivate the trypsin. Pipette cells up and down ~ 5 times with a 10 mL pipette to obtain a single cell suspension, while avoiding frothing of medium. Plate cells into new sterile flasks or plates containing appropriate Cell Culture Medium. Place the cells in a humidified 37 °C incubator with 5% CO <sub>2</sub> . |
| Recommended Cryopreservation<br>Medium | 45% IMDM, 50% FBS, 5% DMSO  |

## **Additional Information**

For the full Technical Manual and protocols, please visit horizondiscovery.com

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